Antioxidant & anticancer activities of isatin (1H-indole-2,3-dione), isolated from the flowers of *Couroupita guianensis* Aubl

Mariappan Premanathan¹, Srinivasan Radhakrishnan², Kumaraasamy Kulinjappar³, Ganesan Singaravelu⁴, Velayutham Thirumalaiarasu⁵, Thangavel Sivakumar⁵ & Kandasamy Kathiresan⁶

¹Department of Biotechnology, Mepco Schlenk Engineering College, Sivakasi, ²Department of Instrumentation & Electroorganic Division, CSIR-Central Electro Chemical Research Institute, Karaikudi, ³Department of Zoology, Thiruvalluvar University, Vellore, ⁴Department of Microbiology, Ayya Nadar Janaki Ammal College, Sivakasi & ⁶Centre of Advanced Study in Marine Biology, Annamalai University, Parangipettai, India

Received July 13, 2010

**Background & objectives:** Derivatives of isatin are known to have cytotoxicity against human carcinoma cell lines. This compound therefore, has a potential to be used as a chemotherapeutic agent against cancer. This study was done to investigate the antioxidant and anticancer activities of isatin, extracted from flower of a folklore medicinal plant *Couroupita guianensis* against human promylocytic leukemia (HL60) cells.

**Methods:** Active fractions demonstrating anticancer and antioxidant activities were isolated from the extracts of shade-dried flowers of *C. guianensis* by bioassay guided fractionation. The free radical scavenging activity was determined using lipid peroxidation assay. Cytotoxicity against human promylocytic leukemia HL60 cells was determined by MTT assay. Apoptotic activity was analyzed by DNA fragmentation and flowcytometry.

**Results:** Isatin isolated from the active fraction showed antioxidant activity with the EC⁵₀ value of 72.80 µg/ml. It also exhibited cytotoxicity against human promylocytic leukemia HL60 cells in dose-dependant manner with the CC⁵₀ value of 2.94 µg/ml. The isatin-treated cells underwent apoptosis and DNA fragmentation. Apoptosis was confirmed by the FACS analysis using FITC-annexin V markers.

**Interpretation & conclusions:** Isatin showed antioxidant activity and was cytotoxic to the HL60 cells due to induction of apoptosis. The isatin can be further evaluated to be used as a prophylactic agent to prevent the free radical-induced cancer and as a chemotherapeutic agent to kill the cancer cells.

**Key words** Anticancer - antioxidant - apoptosis - *Couroupita guianensis* - free radicals - isatin
Isatin (1H-indole-2,3-dione) is an endogenous compound, identified in humans and possesses a wide range of biological activities such as anxiogenic, sedative, anticonvulsant activities and acts as a potent antagonist on atrial natriuretic peptide receptors in vitro1,2. Derivatives of isatin have been reported to have cytotoxicity against human carcinoma cell lines derived from breast, prostate1, human acute lymphoblastic leukemia (MOLT-4)2,3, colon2,3, and lung3. Isatin was first isolated from the fruits of the Cannon ball tree Couroupita guianensis9. This plant species has been used in folklore medicines10,11. However, it is not scientifically validated. We report here the potential of isatin isolated from the flowers of C. guianensis for antioxidant and anticancer activities against human promyelocytic leukemia (HL60) cells.

**Material & Methods**

Couroupita guianensis Aubl. (Lecythidaceae) is an evergreen tree, native to tropical northern South America, southern Caribbean and also India. Its flowers are orange, scarlet and pink in colour, and form large bunches. Floral parts of C. guianensis were collected from Kattalagar kovil with elevation of 100.07 m above sea level, Madurai, Tamil Nadu, India (9° 28’ N and 77° 48’ E). The voucher specimen was identified and deposited in the herbarium of the Centre of Advanced Study in Marine Biology, Parangipettai, Tamil Nadu. The samples were washed, air-dried and powdered.

**Extraction and purification:** Extraction and purification of the floral parts were carried out as described earlier9. Dried, powdered (250 g) floral parts were extracted with CHCl3 using soxhlet extractor and concentrated in a rotary vacuum evaporator. The residue thus obtained was dissolved in CH3Cl2 and passed through a column of silica gel to remove the colouring material. The resulting solution was concentrated and chromatographed on silica gel column (5 x 50 cm, 60-120 mesh) using CH3Cl2 containing slowly increasing amounts of methanol as eluent (flow rate: 3 ml/min). Fractions of 250 ml were collected and monitored by HPLC (Shimadzu, Japan) using shimpack ODS-18 column as stationary phase (4.5 x 120 mm), eluted with methanol-water 80:20 at a flow rate of 1 ml/min. The samples were analysed using a UV detector at a wave length of 295 nm. Elution of column chromatography afforded four major fractions. Each fraction was concentrated using a rotary evaporator and the residue was tested for its biological activity. The third fraction which showed promising activity was analyzed for structural elucidation by infra red (IR), ultra violet (UV), 1H nuclear magnetic resonance (NMR), 13C NMR and gas chromatography-mass spectral technique (GC-MS) spectral techniques.

IR spectra, GC-MS and NMR analysis were made at CSIR-Central Electro Chemical Research Institute, Karaikudi, Tamil Nadu using a Bruker Optik GmbH, make TENSOR 27 spectrometer (Germany), Agilent GC-MS (USA) and Bruker 400 MHz NMR spectrometer (Switzerland), respectively.

**Cytotoxicity assay:** HL60 cells obtained from the National Centre for Cell Science, Pune, India were grown in RPMI1640 medium supplemented with 2 mg/ml sodium bicarbonate, 4.5 mg/ml glucose, 100 µg/ml streptomycin sulphate, 40 µg/ml gentamycin, 100 U/ml penicillin as well as 10 per cent heat inactivated foetal calf serum. An environment of humidified air containing 5 per cent CO2 was maintained at 37°C. Cytotoxicity was determined by MTT assay12,13. Briefly, lipid peroxidation was induced in liposome prepared from egg lecithin by adding 5 µl of 400 mM FeCl3 and 5 µl of 200 mM L-ascorbic acid. To this, different concentrations of the test compound were added. The control was prepared without the treatment of compound. The samples were incubated at 37°C for 60 min. The reaction was inhibited by adding 1 ml of stopping solution which contains 0.25 N HCl, 1.5 per cent trichloroacetic acid, 0.375 per cent thiobarbituric acid. These reaction mixtures were kept in boiling water bath for 15 min, cooled and centrifuged. The absorbance of the resulting solution was measured at 532 nm. The activity was calculated by using the formula: 50% inhibition (EC50) = [(control OD - sample OD) / control OD] x 100.

**DNA fragmentation study:** HL60 cells were incubated with appropriate concentration of the test compound
with their CC₅₀ value. After 48 h DNA was extracted using DNA isolation kit (Genei, Bangalore, India), evaluated on 0.8 per cent agarose gel using ethidium bromide and DNA pattern was documented by gel documentation system (Vilber Lourmet, France)³³.

Apoptosis detection by Annexin V marker: HL60 cells were incubated with appropriate concentration of the test compound with their CC₅₀ value. After 24 h, apoptosis induction was analyzed using the apoptotic, necrotic, and healthy cell quantification kit (Biotium inc., USA) following the manufacturer’s protocol for flow cytometry (FACS caliber, BD Biosciences, USA) assay¹³,¹⁴.

Statistical analysis: Data were analysed using Biostat, Analyst Soft Inc. software using one-way (ANOVA) for statistical significance of the model with post hoc comparisons to test for statistically significant effects.

Results & Discussion

The chemical fractions extracted from flower of the Cannonball tree exhibited promising biological activity only in the third fraction and hence this fraction was analyzed by Fourier Transform Infra Red (FTIR), UV, ¹H NMR, ¹³C NMR, GC-MS and CHN Elemental analysis techniques for its structural elucidation.

FTIR spectrum (Fig. 1) showed stretching bands at 1730 and 1616/cm representing the presence of carbonyl and secondary amino groups, respectively. A peak at a wave number of 3187/cm confirmed the existence of amino functional group. Carbon-hydrogen stretching in aromatic was observed at 3100-3000/cm.

The UV spectrum of the active fraction showed the absorption at a wavelength of 240 and 280 nm in methanol medium. In the ¹H NMR spectrum (Bruker FT-NMR, 400 MHz) four aromatic protons were observed at δ 7-8. ¹³C NMR spectrum indicated eight different carbon atoms. ¹³C NMR (100 MHz, Acetoned-δ): δ 112, 118, 123, 125, 138, 151, 159, 184. Elemental analysis showed: C, 65.26; H, 3.40; N, 9.52. C₈H₅NO₂ requires: C, 65.31; H, 3.45; N, 9.48.

The GC-MS was operated at the oven temperature of 250°C with Quadrupole detector. The spectrum of the isolated compound indicated the presence of molecular ion (m/z = 147[M⁺]). The purified compound was confirmed to be isatin (1H-indole-2,3-dione) and its yield was 3 mg (melting point: 203°C) from 250 g dry powder of flower.

The antioxidant activity of the purified compound was measured by the inhibition of lipid peroxidation. Isatin inhibited the ultrasound induced lipid peroxidation in liposome prepared from egg lecithin. It showed the radical scavenging activity with EC₅₀ of 72.80 µg/ml (Fig. 2). Ascorbic acid was used as positive control with EC₅₀ of 754.14 µm.

The cytotoxicity of isatin was studied in the human promyelocytic leukemia HL60 cells. The 50 per cent cytotoxicity (CC₅₀) values were determined after five days of exposure. Isatin showed cytotoxicity in dose-dependent manner at CC₅₀ of 2.94 µg/ml (Fig. 2); 5-Fluourouracil was used as positive control with CC₅₀ of 0.07 µm.

Studies were performed to determine if isatin-induced cytotoxicity occurred via an apoptotic pathway. The DNA profile of the cells treated with the purified compound along with control cells was analyzed. In control cells, the DNA was not fragmented whereas the cells treated with isatin showed the apoptosis and DNA fragmentation (data not shown). FACS analysis was carried out to confirm the apoptotic activity (Fig. 3).

There are a few reports on the isolation of isatin from the genus *Isatis*⁶, in fruits of the cannon ball tree, *C. guianensis* Aubl⁹ and in secretions from the parotid gland of the *Bufo* frog¹⁷. Various substituted isatins have also been identified in plants¹⁸,¹⁹, fungi²⁰ and marine molluscs²¹.

Isatin inhibited the ultrasonic radiation induced lipid peroxidation in liposomal membrane with EC₅₀ of 72.80 µg/ml. Increase in lipid peroxidation denotes
cytotoxicity and hepatocellular dysfunction in mice. In the present study isatin showed antioxidant activity with EC50 value of 72.80 µg/ml and cytotoxicity in HL60 cells with CC50 of 2.94 µg/ml. We have earlier shown that compounds which exhibit high cytotoxicity have less antioxidant activity.

To interpret the data regarding the mode of cell death after exposure to isatin, it is recommended that a combination of at least three criteria of cell death be evaluated: cell morphology, DNA fragmentation, annexin V binding, and/or caspase activation. Hence, in the present study isatin induced cytotoxicity was determined through an apoptotic pathway. Isatin started the apoptosis process with fragmentation of DNA (data not shown). Cleavage of DNA at the internucleosomal linker sites yielding DNA fragments is regarded as a biochemical hallmark of apoptosis. Apoptosis induced by isatin was confirmed by flow cytometry to further elucidate the extent and causes of apoptosis.

In conclusion, isatin was isolated from the floral parts of cannon ball tree and it exhibited antioxidant activity and cytotoxicity against HL60 cells. Isatin can be further evaluated to be used as prophylactic agent to prevent the free radicals induced cancer and as chemotherapeutic agent to kill the cancer cells.

Acknowledgment

The authors thank the Management, Principal of the Mepco Schlenk Engineering College (M.P) and Ayya Nadar Janaki Ammal College (V.T & T.S) Sivakasi, Tamil Nadu, India for providing the necessary facilities to carry out this work. FACS analysis was done with the help of C. Gowri Priya at TIFAC-Center of Relevance and Excellence in Diabetic Retinopathy, Aravind Medical Research Foundation, Aravind Eye Hospital, Madurai, Tamil Nadu, India.

References

Directorate, Council of Scientific & Industrial Research; 1956.


Reprint requests: Dr M. Premanathan, Department of Biotechnology, Mepco Schlenk Engineering College, Mepco Nagar, Sivakasi 608 502, Tamil Nadu, India e-mail: ma.premanathan@gmail.com